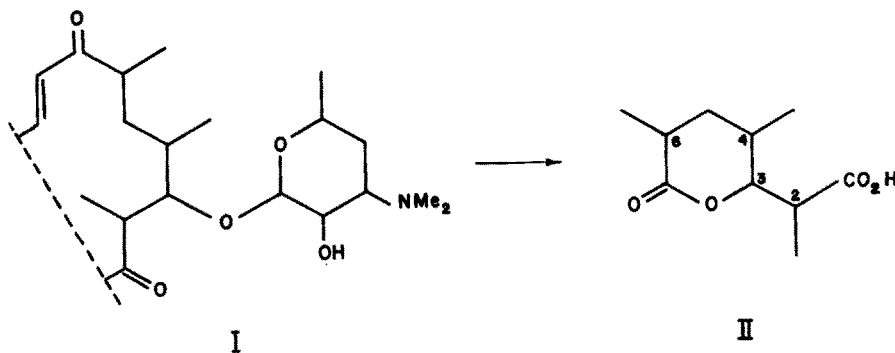


MACROLIDE ANTIBIOTIC STUDIES. XIII*
PARTIAL ABSOLUTE CONFIGURATIONS OF METHMYCIN, NEOMETHMYCIN,
NARBOMYCIN AND PICROMYCIN

R.W. Rickards and R.M. Smith
Research School of Chemistry, Australian National University,
Canberra, A.C.T.

(Received in UK 6 February 1970; accepted for publication 12 February 1970)

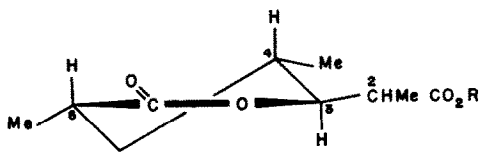
Oxidative degradation of the macrolide antibiotics¹ methmycin², neomethmycin³, narbomycin⁴, and picromycin⁵, all of which contain the structural segment (I), yields the (+)-lactonic acid (II)^{2,6}. This key degradation product was assigned the absolute configuration 4S, 6R by Djerassi and co-workers⁷ and 2S, 3R by Bergel'son and Batrakov⁸, and corresponding stereochemical assignments were then made in the parent macrolides^{7,8}. We show here that the 2S, 3R assignments⁸ in the acid (II), and the resulting assignments⁸ in the antibiotics themselves, should be reversed.



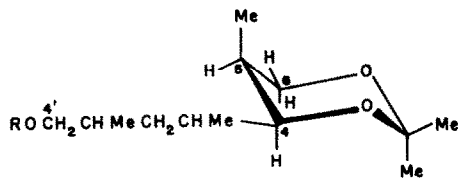
The pmr spectrum (in CDCl_3) of the natural lactonic acid (II) showed H_3 as a double doublet (J 2.3, 10.0 Hz) centred at τ 5.45, and a double quartet (J_d 2.3, J_q 7.5 Hz) at τ 7.27, which was confirmed as H_2 by irradiation at τ 5.45 when it collapsed to a quartet (J 7.5 Hz).

* Part XII, D.G. Manwaring, R.W. Rickards and B.T. Golding, submitted for publication.

Irradiation of H_2 collapsed the H_3 signal to a doublet (J 10.0 Hz) and a $CHCH_3$ doublet at τ 8.78 (J 7.5 Hz) to a singlet. Thus $J_{3,4}$ must be 10.0 Hz, whilst $J_{2,3}$ must be 2.3 Hz. Similar pmr results were obtained for the lactonic acid methyl ester, in which H_3 resonated as a doublet at τ 5.50 with $J_{3,4}$ 10.0 Hz and $J_{2,3}$ 2.3 Hz, and H_2 as a double quartet, τ 7.30 (J_d 2.3, J_q 7.0 Hz). The magnitude of $J_{3,4}$ in these compounds necessitates⁹ that these vicinal protons are anti-periplanar¹⁰, and in conjunction with the previous 4S assignment⁷ leads to the S configuration for C-3. The most likely conformation for the lactones is the half-chair form¹¹ (III; R = H or Me), although a half-boat form¹¹ is not excluded on pmr grounds.



III



IV

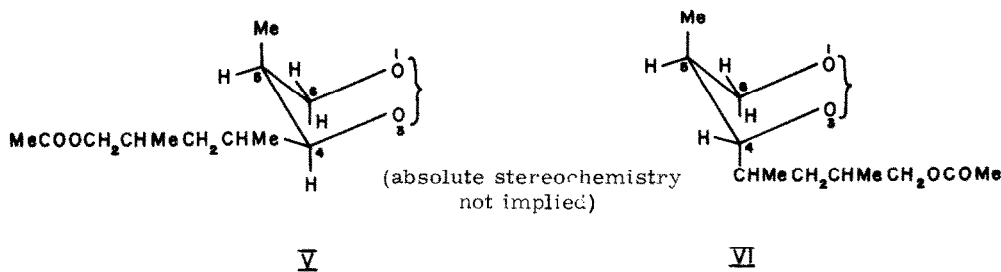
In order to relate the stereochemistry of C-2 to C-3 in the acid (III; R = H), the ester (III; R = Me) was reduced with lithium aluminium hydride to the triol⁸, which with acetone and *p*-toluenesulphonic acid gave an hydroxyketal⁸. The 1,3-dioxan structure (IV; R = H) of this ketal follows from major mass spectroscopic fragmentations of the acetyl derivative (IV; R = MeCO) to give m/e 257 and 129, representing $(M - Me)^+$ and $(M - CHMeCH_2CHMeCH_2OCOMe)^+$ ions.

Pmr spectra of the acetylated 1,3-dioxan (IV; R = MeCO) showed resolved absorption due to five protons between τ 5.7 - 6.8, which could be interpreted with the aid of double irradiation and solvent shifts. The 4'-methylene protons appeared (in C_6D_6) at lowest field as the AB portion of an ABX system, double doublets too close in chemical shift for first order analysis. Three further double doublets centred at τ 6.25 (J 11.5, 2.5 Hz), 6.60 (J 11.5, 1.5 Hz) and 6.81 (J 8.9, 2.3 Hz) represent respectively the two mutually-coupled C-6 methylene protons* and the C-4 methine proton of the 1,3-dioxan ring. Irradiation at the H_5 frequency (τ 8.82) causes collapse of all three resonances to simple doublets with loss of the smaller coupling in each case. It follows from the Karplus dihedral angle relationship⁹, coupled with the fact

* These protons form the AB portion of an ABX system, the first order analysis of which was validated by an ABX computation of line positions and intensities.

that the system C-4,5,6 is part of a 6-membered ring, that H_5 is syn-clinal¹⁰ to both H_6 protons and also to H_4 . Two partial approximate conformations (V) and (VI) of the dioxan ring result. (VI) with the C-2 gem-dimethyl group added would involve severe axial-type interactions^{12,13} and is not a possible preferred conformation. (V) remains, from which it follows that H_4 and H_5 are cis-related, and that the absolute configuration of C-5 is S.

These conclusions are valid regardless of the relative position of C-2, i.e., regardless of the overall conformation of the 1,3-dioxan ring. However, of the various conformations possible, the chair form (IV) is likely to be preferred, since boat and skew-boat forms will be destabilised by 2,5- and 2,4-alkyl-alkyl interactions^{12,13}. This chair conformation (IV) and the embodied relative configurational assignments at C-4 and C-5 are confirmed by published chemical shift¹⁴ and coupling constant^{12,14} data, which also permit assignment of the τ 6.25 and 6.60 resonances to the equatorial and axial protons respectively at C-6. Thus in a series of substituted 1,3-dioxans, H_{4eq} and H_{6eq} resonated in the range τ 5.86 - 6.24, H_{4ax} and H_{6ax} at higher field between τ 6.44 - 6.96, whilst H_{5eq} resonated at τ 8.80 - 8.81 with H_{5ax} to lower field τ 7.97 - 8.09. $J_{4ax,5eq(6ax,5eq)}$ ranged between 2.5 - 4.4 Hz, $J_{4eq,5eq(6eq,5eq)}$ 1.2 - 1.9 Hz, $J_{4eq,5ax(6eq,5ax)}$ 3.3 - 6.6 Hz, $J_{4ax,5ax(6ax,5ax)}$ 9.0 - 13 Hz, and $J_{6eq,6ax}$ 10.1 - 11.3 Hz.



The complete absolute stereochemistry of the natural lactonic acid (II) can now be designated as 2R, 3S, 4S⁷, 6R⁷, thus reversing Bergel'son and Batrakov's assignments^{*8} at C-2 and C-3. Extension of these assignments to the parent antibiotics defines the partial absolute configuration of methymycin² and neomethymycin³ as 2R, 3S, 4S⁷, 6R⁷, and of narbomycin⁴ and picromycin⁵ as 4R, 5S, 6S^{7,5}, 8R^{7,5}. Allowing for the revision⁵ of picromycin's structure, these conclusions are in complete agreement with the predictions of Celmer's

* These authors have apparently proven the relative stereochemistry of a racemic diastereoisomer of the natural acid (II). Their error probably stems from an incorrect identification, reportedly¹⁵ based only on infrared spectra, of this diastereoisomer as the racemate of the natural acid.

configurational model¹⁶ for macrolide antibiotics.

Acknowledgements: We thank Mr C. Arandjelovic for pmr spectra, and Dr R. Bramley for helpful discussion.

REFERENCES

1. R.B. Woodward, Festschrift Arthur Stoll, p.524, Birkhauser, Basel (1957), and Angew. Chem. 69, 50 (1957); M. Berry, Q. Rev. chem. Soc. 17, 343 (1963).
2. C. Djerassi and J.A. Zderic, J. Am. chem. Soc. 78, 2907, 6390 (1956); and references therein.
3. C. Djerassi and O. Halpern, J. Am. chem. Soc. 79, 2022 (1957), and Tetrahedron 3, 255 (1958).
4. V. Prelog, A.M. Gold, G. Talbot and A. Zamojski, Helv. chim. Acta 45, 4 (1962); and references therein.
5. R.W. Rickards, R.M. Smith and J. Majer, Chem. Comm., 1049 (1968); H. Muxfeldt, S. Shrader, P. Hansen and H. Brockmann, J. Am. chem. Soc. 90, 4748 (1968); and references therein.
6. R. Anliker, D. Dvornik, K. Gubler, H. Heusser and V. Prelog, Helv. chim. Acta 39, 1785 (1956).
7. C. Djerassi and O. Halpern, J. Am. chem. Soc. 79, 3926 (1957); C. Djerassi, O. Halpern, D.I. Wilkinson and E.J. Eisenbraun, Tetrahedron 4, 369 (1958).
8. L.D. Bergel'son and S.G. Batrakov, Bull. Acad. Sci. U.S.S.R. Div. chem. Sci., 1982 (1966).
9. M. Karplus, J. Am. chem. Soc. 85, 2870 (1963); J.W. Emsley, J. Feeney and L.H. Sutcliffe, High Resolution Nuclear Magnetic Resonance Spectroscopy 1, p.166, and 2, p.678, Pergamon, Oxford (1965); H. Conroy, Advances in Organic Chemistry 2, p.265, ed. R.A. Raphael, E.C. Taylor and H. Wynberg, Interscience, New York (1960); and references therein.
10. W. Klyne and V. Prelog, Experientia 16, 521 (1960).
11. A.F. Beecham, Tetrahedron Lett., 3591 (1968); H. Wolf, Tetrahedron Lett., 5151 (1966); K.K. Cheung, K.H. Overton and G.A. Sim, Chem. Comm., 634 (1965); and references therein.
12. J.E. Anderson, F.G. Riddell and M.J.T. Robinson, Tetrahedron Lett., 2017 (1967).
13. K. Pihlaja, Acta chem. scand. 22, 716 (1968).
14. M. Anteunis, D. Tavernier and F. Borremans, Bull. Soc. chim. Belg. 75, 396 (1966).
15. L.D. Bergel'son and S.G. Batrakov, Bull. Acad. Sci. U.S.S.R. Div. chem. Sci., 1259 (1963).
16. W.D. Celmer, J. Am. chem. Soc. 87, 1801 (1965); Antimicrobial Agents and Chemotherapy 1965, p.144, ed. G.L. Hobby, Am. Soc. Microbiol., Ann Arbor (1966); Biogenesis of Antibiotic Substances, p.99, ed. Z. Vanek and Z. Hostalek, Czechoslovak Academy of Sciences, Prague (1965); J. Am. chem. Soc. 88, 5028 (1966).